

Kinesin-8 molecular motors: putting the brakes on chromosome oscillations

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Recent studies suggest that the human Kinesin-8 molecular motor Kif18A has a role in chromosome congression. Specifically, these studies find that Kif18A promotes chromosome congression by attenuating chromosome oscillation magnitudes. Together with recent modeling work, *in vitro* studies, and the analysis of *in vivo* yeast data, these reports reveal how Kinesin-8 molecular motors might control chromosome oscillation amplitudes by spatially regulating the dynamic instability of microtubule plus-ends within the mitotic spindle.

Introduction

The mitotic spindle is a microtubule-based structure that controls the proper segregation of chromosomes during mitosis [1]. Within the mitotic spindle, microtubules coordinate the congression of chromosomes to the metaphase plate before their ultimate segregation during anaphase. Although chromosomes generally align at the spindle equator, this alignment is dynamic, with chromosomes oscillating about this position throughout metaphase. Because the microtubules that coordinate chromosome congression are dynamic – growing and shortening through a behavior termed ‘dynamic instability’ [2] – it is likely that chromosome oscillations reflect the growth and shortening dynamics at microtubule ends. Therefore, a central issue in the study of mitosis is describing how the dynamics of microtubule ends are regulated spatially within the mitotic spindle such that chromosomes are congressed to the spindle equator.

Microtubules are composed of $\alpha\beta$ -tubulin heterodimers that are stacked end-on, typically into 13 individual protofilaments. Growth and shortening at microtubule ends takes place primarily at the β -tubulin exposed ‘plus-end’ of microtubules, through the addition of GTP–tubulin heterodimers [3]. After their addition, lattice-bound GTP–tubulin subunits are hydrolyzed into GDP–tubulin subunits through nucleotide exchange on the β subunit. Importantly, GDP–tubulin subunits exposed at the tips of microtubules are unstable owing to a conformational change in the tubulin. This promotes the curling and peeling of individual microtubule protofilaments, thus stretching and destabilizing bonds between adjacent tubulin subunits. Stochastic switching between stable, straight, growing GTP–tubulin-capped microtubules and unstable, peeling, shortening GDP–tubulin microtubules forms the basis of microtubule dynamic instability. The frequency at

which microtubules switch from the shortening to the growing state is termed the ‘rescue’ frequency; and the frequency at which the reverse switch – from growing to shortening – occurs is termed the ‘catastrophe’ frequency.

Microtubule-associated proteins (MAPs) associate with the microtubule lattice and can modulate the dynamics of microtubule ends. Members of the Kinesin-8 family of molecular motors are MAPs that have been classified as catastrophe promoters based on the observation that motor deletion mutants exhibit abnormally long and stable microtubules [4–7]. A recent *in vitro* study with GTP–tubulin-stabilized microtubules [or guanylyl-(α , β)-methylene-diphosphonate (GMPCPP) microtubules] suggested that the yeast Kinesin-8 motor Kip3p promotes shortening of microtubule plus-ends in a length-dependent manner [8]. In this study, GMPCPP microtubules were used because GMPCPP mimics the GTP ‘cap’ found at the ends of growing microtubules. Here, the presence of Kip3p at moderate concentrations promoted rapid depolymerization of longer microtubules and promoted slower depolymerization of shorter microtubules (Table 1) [8]. Because Kip3p is a molecular motor, this result probably originates from the ability of Kip3p to specifically motor towards the plus ends of microtubules, concentrating in a length-dependent manner on the plus ends of longer microtubules. Upon arrival at the plus end, the motor then destabilizes interactions between neighboring tubulin dimers. Because longer microtubules have more potential Kip3p-binding sites, they will tend to attract more motors, so that there will be a higher concentration of microtubule-destabilizing Kip3p motors on the ends of longer microtubules, and a lower concentration at the ends of shorter microtubules. A recent study by Mayr and co-workers [9] described a similar property for the human Kinesin-8 motor Kif18A (Table 1).

Kinesin-8 motors are localized to microtubules in the mitotic spindle [10], and deletion of Kinesin-8 motors prevents proper chromosome congression in some organisms [9,11–14]. Thus, in this review we discuss how the principles revealed through these *in vitro* studies might provide a mechanism by which chromosome oscillations and the congression of chromosomes to the spindle equator take place during mitosis.

Kinesin-8 motors and the mitotic spindle

Recently, Stumpff et al. [15] addressed this question by carefully quantifying kinetochore oscillations in mitotic HeLa cells that had varying expression levels of the Kinesin-8 molecular motor Kif18A. Surprisingly, the *in vivo*

Table 1. Effect of Kinesin-8 on microtubule-shortening rate and catastrophe frequency

| Parameter | Experiment type and organism | With Kinesin-8 ^a | Without Kinesin-8 ^b |
|---|--|----------------------------------|----------------------------------|
| Microtubule-shortening rate ($\mu\text{m}/\text{min}$) | <i>In vivo</i> : budding yeast astral microtubules | 1.91 ± 0.69 [16] | 2.82 ± 0.95 [16] |
| | <i>In vivo</i> , human (HeLa cells) | 1.92 ± 0.06 [15] (metaphase) | 2.80 ± 0.08 [15] (metaphase) |
| | | 1.68 ± 0.08 [15] (anaphase) | 2.12 ± 0.14 [15] (anaphase) |
| | <i>In vitro</i> , GMPCPP microtubules | ~ 0.10 [16] (50 nM Kip3p) | ~ 0.00 [16] |
| | | ~ 0.50 [8] (3.3 nM Kip3p) | ~ 0.00 [8] |
| Microtubule catastrophe frequency (min^{-1}) | <i>In vitro</i> , GMPCPP microtubules | ~ 1.25 [9] (100 nM Kif18A) | ~ 0.00 [9] |
| | <i>In vivo</i> , budding yeast astral microtubules | 0.48 [16] | 0.27 [16] |

^aKinesin-8 molecular motors are present in the assay, either endogenously as in the *in vivo* studies, or added to the assay in the noted concentrations as in the *in vitro* studies.

^bKinesin-8 molecular motors are either deleted or knocked down as in the *in vivo* studies, or absent as in the *in vitro* studies.

effect of Kinesin-8 molecular motors on microtubule depolymerization rates appears to be the opposite of the reported *in vitro* effect on GMPCPP-stabilized microtubules. Although the presence of Kinesin-8 motors results in more rapid *in vitro* depolymerization of longer GMPCPP-stabilized microtubules, the presence of Kif18A *in vivo* appears to slow the rate of microtubule-shortening during both metaphase and anaphase (Table 1) [15]. This result is consistent with recent *in vivo* measurements of yeast cytoplasmic microtubule dynamics, in which shortening rates were slowed by $\sim 48\%$ in the presence of Kip3p (Table 1) [16]. Mayr *et al.* [9] reported slower *in vivo* chromosome oscillation velocities with Kif18A depletion. However, these velocities were probably underestimated, because the observed frame-rate (0.91 frames/ min^9) was similar to the kinetochore-switching rate (in cells depleted of Kif18A, the switching rate was ~ 1.23 switches/ min^{15}). Thus, at least in part, the loss of chromosome congression in human HeLa cells depleted of Kif18A is the result of larger chromosome oscillation amplitudes that originate from rapid microtubule depolymerization in the absence of Kif18A.

Depolymerization at the microtubule plus-end

It is interesting to consider how these opposing effects of Kinesin-8 on *in vitro* GMPCPP microtubule depolymerization rates and on *in vivo* GDP-tubulin microtubule depolymerization rates might arise. Given that stable GMPCPP microtubules are composed primarily of GTP-tubulin subunits, *in vitro* GMPCPP microtubule protofilaments prefer to be relatively straight. This means that the strain between the motor domains of the plus-end-directed Kinesin-8 molecular motor might force outward protofilament curling (Figure 1a, upper panel). This curling would destabilize the lateral bonds with neighboring tubulin subunits, which would then increase the dissociation rate of the most distal tubulin subunit, provided that the lost lateral-bond energy and the gained curling-energy together exceed the motor-cross-linking energy. Based on our previous modeling of microtubule disassembly [17,18], we estimate that the lost lateral-bond energy in the presence of the motor is $\sim +5$ $k_B T$ (this is for a single bond, where 1 $k_B T$ /single bond = 0.6 kcal/mol of bonds; note that a high-affinity bond, $K_D \sim 1$ nM, has a standard Gibbs free-energy change of -21 $k_B T$, and that a moderate affinity bond, $K_D \sim 1$ μM , has an energy change of -14 $k_B T$). Furthermore, we estimate that the curling energy penalty is $\sim +3$ $k_B T$, for a total of $+8$ $k_B T$ of unfavorable energy. Given that this is less than the Gibbs free-energy of GTP hydrolysis (i.e. ~ -20 $k_B T$), it is feasible for the motor to generate such

strains. By comparison, the longitudinal cross-linking energy of XMAP215 – in this case, it is considered to be a MAP that can cross-link tubulin dimers in a manner similar to Kif18A – was estimated to be ~ -4 $k_B T^{17}$. Thus, there is a possible net increase of $\sim +4$ $k_B T$ of unfavorable energy due to outward protofilament-curling generated from strain between the motor-walking domains. This would increase the tubulin dissociation rate by more than 50-fold. A similar Kinesin-8-mediated mechanism would then presumably apply to growing microtubules *in vivo* with a stabilizing GTP-tubulin cap. Curling of GTP-tubulin subunits at growing microtubule tips might promote removal of the stable GTP-tubulin cap and this in turn would initiate microtubule catastrophe by exposing unstable GDP-tubulin subunits at the microtubule tip. The difference between GMPCPP-stabilized microtubules *in vitro* and GDP-tubulin microtubules *in vivo* is that, once uncapped, depolymerization of GDP-tubulin microtubules then proceeds spontaneously through the curling and peeling of unstable GDP-tubulin protofilaments. In this case, Kinesin-8 motors might act passively to cross-link and stabilize the bonds between neighboring tubulin subunits (Figure 1a, lower panel), thereby slowing the *in vivo* depolymerization rate. Thus, it might be that the microtubule-destabilizing effect of Kinesin-8 is manifested *in vivo* by an increase in microtubule catastrophe frequency – as measured recently in experiments with cytoplasmic microtubules in yeast (Table 1) [16] – even though Kinesin-8 motors actually slow the microtubule-shortening rate *in vivo*. This behavior is consistent with the observation that the *Drosophila* Kinesin-8 motor Klp67A tends to destabilize dynamic microtubules during metaphase, while stabilizing the formation of the central spindle during anaphase. These effects might be caused by increased microtubule catastrophe during metaphase with a reduced inter-polar microtubule depolymerization rate during anaphase [4]. It would be interesting to test the effect of Kinesin-8 on dynamic GTP-tubulin microtubule depolymerization rates *in vitro*, in an effort to determine whether these contrasting behaviors can be observed.

Kinesin-8 mediated regulation of chromosome oscillations

As shown by Stumpff *et al.* [15], the increased microtubule-shortening velocity is a major factor in the increased kinetochore oscillation amplitude in Kif18A-depleted cells. In these cells, it is likely that shortening microtubules drive pole-ward motion of leading kinetochores [19]. As described above, Kif18A might also promote the length-dependent catastrophe of longer microtubules [8,9]. The

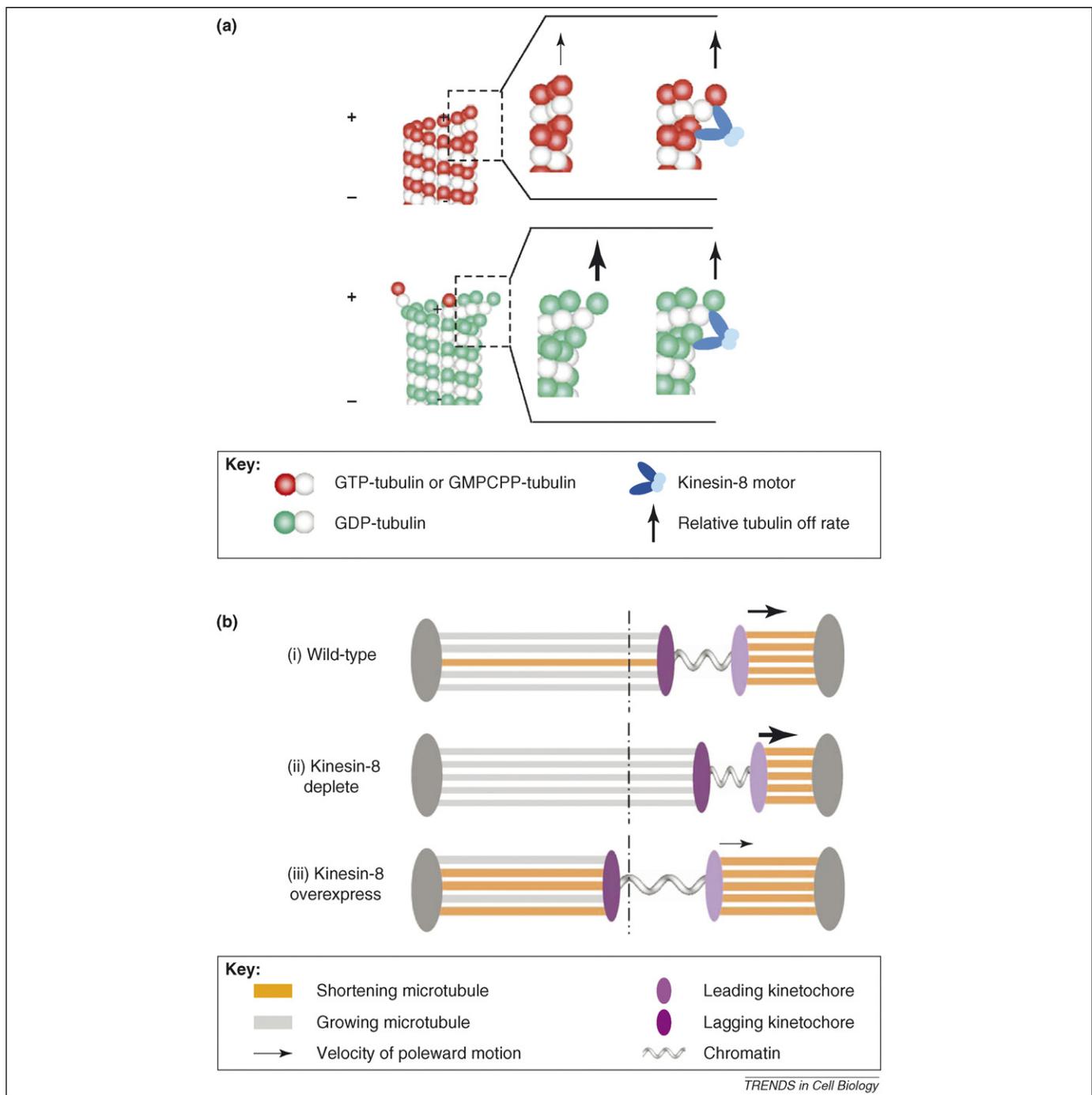


Figure 1. Kinesin-8 motors and microtubules. **(a)** *In vitro* microtubule-shortening rates are higher in the presence of Kinesin-8 molecular motors, whereas the opposite is observed *in vivo*. Stable GMPCPP microtubules used in the *in vitro* experiments (upper panel) might be more sensitive to mechanical tip destabilization through Kinesin-8-mediated protofilament kinking, whereas GDP-tubulin microtubules *in vivo* (lower panel) show rapid, spontaneous depolymerization; thus Kinesin-8 molecular motors might act to passively slow the natural depolymerization process (see text for more quantitative details). **(b)** A model for Kif18A-mediated kinetochore oscillation amplitude. (i) In wild type spindles, longer microtubules attached to lagging kinetochores are selectively targeted by Kif18A for catastrophe and subsequent depolymerization, decreasing coordination in the polymerization state and slowing the velocity of pole-ward motion (dotted line represents the spindle equator). (ii) In Kif18A depletion, longer, polymerizing microtubules are not targeted for catastrophe and depolymerization. In addition, rapid microtubule-shortening results in faster pole-ward movement of the leading kinetochore. (iii) Conversely, overexpression of Kif18A might result in increased catastrophe of microtubules attached to the lagging kinetochore, decreasing the coordination in polymerization state. Combined with a slower microtubule depolymerization rate, pole-ward kinetochore motion is slowed, decreasing oscillation amplitude, and resulting in increased tension between sister kinetochores.

promotion of microtubule plus-end catastrophe based on the length of the microtubule might provide an efficient mechanism for establishing a spatial gradient in spindle microtubule dynamics [20] that would regulate microtubule lengths and thus promote chromosome congression during mitosis [21]. Recent modeling studies provide a framework for integrating collective microtubule dynamics

to better understand their role in chromosome oscillations and congression [22–24]. Specifically, modeling work by Civelekoglu-Scholey *et al.* [22] predicts that, for chromosome oscillations to occur, the polymerization state of microtubule ends must be at least partially synchronized, and that pole-ward chromosome motion is dependent on depolymerization of microtubules attached to the leading

kinetochore (Figure 1b(i)). Thus, the attenuation of the microtubule depolymerization rate in wild type spindles would be predicted to reduce oscillation amplitude (Figure 1b(i)). In addition, microtubule catastrophe promotion, as mediated by Kif18A, would be predicted to exert its effect primarily on the longer microtubules attached to lagging kinetochores (Figure 1b(i)), increasing tension between sister kinetochores and, through reduced synchronization of the polymerization state, limiting oscillation amplitude. As shown in Figure 1b(ii), depletion of Kif18A would thus increase shortening velocity for microtubules attached to the leading kinetochore, and, through a lowered catastrophe frequency for the longer microtubules attached to the lagging kinetochores, increase the synchronization of the polymerization state. Overexpression of Kif18A would have the opposite effect, as shown in Figure 1b(iii). This model predicts that Kif18A accumulates preferentially on longer microtubules attached to lagging kinetochores, which is consistent with the asymmetric distribution of Kif18A on sister kinetochores, as was observed by Stumpff *et al.* [15]. Although a more complete description of kinetochore directional instability probably involves both the coordination of microtubule plus-end polymerization state and the dynamic turnover of microtubule-kinetochore attachments [22–25], the results reported by Stumpff *et al.* [15] suggest that Kinesin-8 regulates microtubule dynamics to limit kinetochore oscillation amplitude and increase tension between sister kinetochores.

Conclusions

Although the recent *in vitro* and *in vivo* Kinesin-8 studies shed significant light on the contribution that this molecular motor makes to mitotic chromosome congression, open questions remain. For example, in budding yeast, in which each kinetochore is associated with a single microtubule, depletion of Kinesin-8 has no apparent effect on proper kinetochore congression [10]. In addition, as reported by Stumpff *et al.* [15], the increase in kinetochore oscillation amplitude in HeLa cells is limited to two-fold, even with >90% depletion of Kif18A. This suggests that additional mechanisms, such as polar ejection forces, contribute significantly to the oscillation and centering of chromosomes during mitosis.

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